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Biochemical Changes Induced by Cartap Hydrochloride (50% SP), Carbamate Insecticide in Freshwater Fish *Cirrhinus mrigala* (Hamilton, 1822)

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ABSTRACT

The freshwater fish *Cirrhinus mrigala* was exposed to Cartap hydrochloride (50% SP) for 24, 48, 72 and 96 h. The LC ₅₀ values were found to be 0.436, 0.419, 0.394 and 0.376 mg⁻¹ in static method and 0.399, 0.371, 0.361 and 0.339 mg.L⁻¹ in continuous flow-through system. The static LC₅₀ values are higher than the continuous flow-through method. The LC₅₀ values showed a decreasing trend with an increase in time of exposure in both the methods. The decrease was more in a continuous flow-through method than in the static method. The fish were exposed to sub-lethal (1/10th of 96 h LC₅₀ value 0.0376 mg.L⁻¹) and lethal (96 h LC₅₀ value 0.376 mg.L⁻¹) concentrations of the pesticide for 24 and 96 hours to study the alterations in glycogen, total proteins and nucleic acids (DNA & RNA) contents of various than in sub-lethal concentrations. From the present study, it can be concluded that Cartap hydrochloride caused a decline in the glycogen, total protein and nucleic acids (DNA, RNA) content in *Cirrhinus mrigala* and the changes are more pronounced in lethal exposure than in sub-lethal exposure.

INTRODUCTION

Indiscriminate use of pesticides is one of the main reasons for the pollution of aquatic ecosystems. These toxic pesticides are causing deleterious effects on aquatic organisms. They are causing stress to aquatic organisms which are reflected as biochemical changes in their body (Mayers 1977). Fish acts as a bioindicator species and can be used for monitoring of water pollution as they accumulate the contaminants from polluted water and diet (Chaudary & Jabeen 2011, Kafilzadeh et al. 2012). Fish accumulate these pollutants directly or indirectly from polluted waters and food chain (Jabeen et al. 2016, Chaudary & Jabeen 2011). Carbamates are extensively used water-soluble pesticides in agricultural practices. In India, Cartap hydrochloride is a Carbamate pesticide which is considered as nereistoxin analog is extensively used in rice and sugarcane crops to control pests. The present investigation is aimed to study the toxic effects of Cartap hydrochloride in sub-lethal and lethal concentrations at 24 and 96 hours of exposure period on glycogen, total protein, DNA and RNA contents of freshwater fish Cirrhinus mrigala (Hamilton 1822).

MATERIALS AND METHODS

The fingerlings of the test fish Cirrhinus mrigala size $6-8 \pm$ $\frac{1}{2}$ cm and weight 6-7 ± $\frac{1}{2}$ g were procured from local fish hatcheries of Nandivelugu, Tenali Mandal, Guntur district, Andhra Pradesh. The fish were acclimated at $(28 \pm 2^{\circ}C)$) in the laboratory conditions for two weeks. All the precautions laid down by APHA et al. (1998) were followed. During the acclimation period, the fish were fed with rice bran and groundnut cake. One day before the experimentation feeding was stopped. Cartap hydrochloride (50% SP) commercial grade was purchased from Mangalagiri, Guntur District. The stock solution was prepared with water as a solvent. The acclimatized fish were exposed to static sub-lethal $(0.376 \text{ mg.L}^{-1})$ and lethal concentrations (3.76 mg.L^{-1}) of Cartap hydrochloride (50% SP) for 24 and 96 h. The hydrographical properties of water were estimated by the modified method followed by Golterman & Claimo (1969) method. Finney's probit analysis (Finney 1971) as reported by Roberts % Boyce (1972) was followed to calculate the LC_{50} value. The 95% confidence limits of the LC_{50} values for each test were also calculated for different time periods by using SPSS software. At the end of the exposure periods, the tissues like gill, brain, liver, kidney and muscle were taken out from exposed and control fish and processed for the estimation of glycogen, total proteins and nucleic acids (DNA & RNA). Glycogen was estimated by the method of Kemp et al. (1954), total protein by Lowry et al. (1951) and DNA and RNA by the methods of Searchy & Maclinnis (1970a & 1970b).

The data obtained in the present work were expressed as means of four observations \pm SD (standard deviation) and were statistically analysed using student "t" test (Pillai & Sinha 1968) to compare means of treated data against their controls and the result was considered significant at (P < 0.05) level.

RESULTS AND DISCUSSION

Glycogen, total proteins and nucleic acids (DNA & RNA) decreased in various tissues, viz. gill, brain, liver, kidney and muscle of *Cirrhinus mrigala* exposed to sub-lethal and lethal concentrations of cartap hydrochloride for 24 and 96 hours were graphically represented in Fig. 1 to Fig. 8. In exposed fish per cent decrease is more apparent in lethal concentrations than at sub-lethal concentrations.

Glycogen

The changes in glycogen content observed in the various tissues of *Cirrhinus mrigala* after the Cartap hydrochloride exposure along with the control are graphically represented in Fig. 1 and Fig. 2.

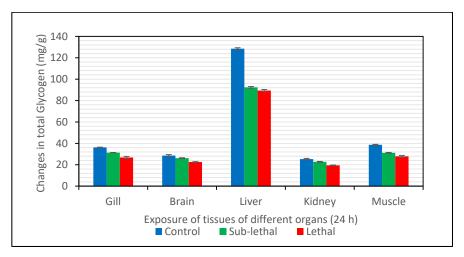


Fig.1: Changes in the glycogen content (mg/g wet weight of the tissue) in the tissue of fish *Cirrhinus mrigala* on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50%SP) for 24 hours.

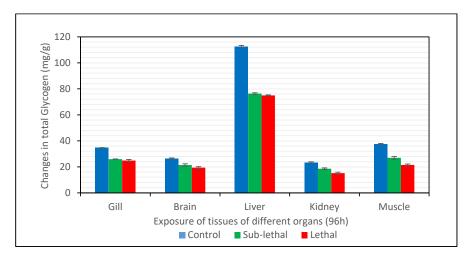


Fig. 2: Changes in the glycogen content (mg/g wet weight of the tissue) in the tissue of fish *Cirrhinus mrigala* on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50% SP) for 96 hours.

In the tissues of control fish, *Cirrhinus mrigala* glycogen content was in the order of:

Liver > Muscle > Gill > Brain > Kidney

Under exposure to sub-lethal and lethal concentrations of Cartap hydrochloride for 24 and 96 hours, the amount of glycogen was found to decrease in all the tissue of *Cirrhinus mrigala*. The lyotropic gradation series in terms of per cent decrement at 24 h and 96 h exposure was:

Sub-lethal -24 h: Liver > Muscle > Gill > Kidney > Brain Lethal -24 h: Liver > Muscle > Gill > Kidney > Brain Sub-lethal 96 h: Liver > Muscle > Gill > Kidney > Brain Lethal-96 h: Muscle > Kidney > Liver > Gill > Brain

Exposure of *Cirrhinus mrigala* to Cartap hydrochloride for 24 hours caused a maximum decrease of glycogen in the liver (sub-lethal 28.117, lethal 30.453). For 96 h maximum decrease was found in the liver (sub-lethal 32.073) and muscle (lethal 42.570). In the present study under Cartap hydrochloride 24 h sub-lethal and lethal exposure minimum percentage of depletion was in the brain (8.595) and (20.96). For 96 h exposure minimum depletion was found in the brain (sub-lethal 18.056, lethal 25.707).

Srivastava & Singh (2013) observed reduction in glycogen content in different tissues of freshwater fish Clarius ba*trachus* exposed to 80% of LC_{50} (22.87 mg.L⁻¹) of Mancozeb at different time intervals of 24 and 96 h. Sastry et al. (1982) reported decreased glycogen content of liver and muscles decreased when Channa punctatus was exposed to sub-lethal concentration of the carbamate pesticide, Sevin (1.05 mg.L^{-1}) for 15, 30 and 60 days. Veeraiah et al. (2013a) observed that exposure to sub-lethal and lethal concentrations of cadmium chloride in the fish Cirrhinus mrigala for 96 h caused changes in the total glycogen level which may be attributed to toxic stress, resulting in the disruption of enzymes associated with carbohydrate metabolism. Bantu & Rathnamma (2013) reported that there is a decrease in the amount of glycogen in the fish Labeo rohita exposed to sub-lethal and lethal concentrations of Dimethoate for 8 days. Priya et al. (2013) observed depletion of glycogen in the liver of freshwater teleost Channa punctatus (Bloch) when exposed to various concentrations of Imidacloprid (0.002 ppm, 0.00 ppm, 0.006 ppm, 0.008 ppm and 0.01 ppm) for 96 h suggesting the possibility of an alter from aerobic to anaerobic mode of energy metabolism of the liver. Dhanalakshmi (2013) noticed decrement in the tissue glycogen concentration in fish Cirrhinus mrigala when exposed to 0.25 ppm concentration of the metal chromium sulphate for 24, 48, 72 h and 10, 20 and 30 days which may be due to its enhanced utilization, since glycogen forms the immediate source of energy to meet energy demands under metallic stress caused by test toxicant.

Tataji & Kumar (2016) reported a decline in glycogen content of freshwater fish Channa punctatus exposed for 8 days to 1/5th of LC₅₀ 96 hours of both Butachlor technical grade and Machete (50% EC), i.e. 32 ppb and 71.2 ppb for both technical and 50% EC respectively. Reduction in glycogen content was also noticed by Naik et al. (2016) in all the tissues of Labeo rohita when exposed to sub-lethal concentrations of cypermethrin for 1, 2 and 3 weeks. Exposure of sub-lethal doses (40% and 80 % of LC₅₀ of 24 h) of glyphosate for 24 or 96 h against the freshwater non-target fish Channa punctatus caused significant (P < 0.05) alteration in biochemical parameters in liver and muscle tissues of the fish Channa punctatus (Bloch) was reported by Singh et al. (2017). Veeraiah et al. (2018) observed that exposure to lethal and sub-lethal concentrations of cyhalothrin 2.5% EC, in the fish Ctenopharyngdon idellus for 24 and 96 hours caused a decrease in glycogen content in all tissues.

According to Dezwaan & Zandee (1973) depletion of glycogen in tissues may be due to direct utilization of the compound for energy generation, a demand caused by pesticide-induced hypoxia. Under hypoxia condition, the fish derives its energy from anaerobic breakdown of glucose which is available to the cell by increased glycogenolysis (Chandravathy & Reddy 1996, Rajamannar & Manohar 1998, Rajamanickam 1992). Reduction in glycogen is probably due to its more rapid break down for energy requirement of fish (Muley et al. 1996).

Liver suggested as an organ for detoxification. During exposure to Cartap hydrochloride exposure fishes came under stress condition and need more energy to cope with the toxicants. glycogen serves as reserve material. It is utilized when the body came under stress condition. Depletion of glycogen in liver and tissues may be due to increment in the glycolysis pathway. During stress conditions, the glycogen reserves depleted to meet energy demand (Rawat et al. 2002). Fall in glycogen levels indicates its rapid utilization to meet the enhanced energy demands intoxicant treated animals through glycolysis or hexose monophosphate pathway as observed by Cappon & Nicholas (1975). The above findings support the alterations of glycogen in the present study.

Proteins

The changes in protein content observed in the various tissues of *Cirrhinus mrigala* after Cartap hydrochloride exposure along with the control was graphically represented in Fig. 3 and Fig. 4.

The Protein content in different tissues in control fish *Cirrhinus mrigala* was in the order of:

Muscle > Liver > Brain > Kidney > Gill

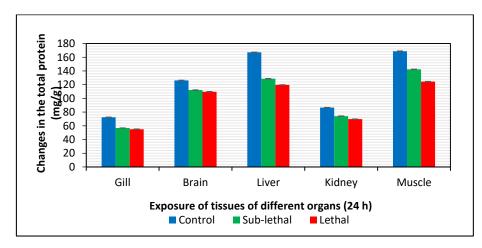


Fig. 3: Changes in protein content (mg/g wet wt of the tissue) in different tissues of fish Cirrhinus mrigala (Hamilton) on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50% SP) for 24 hours.

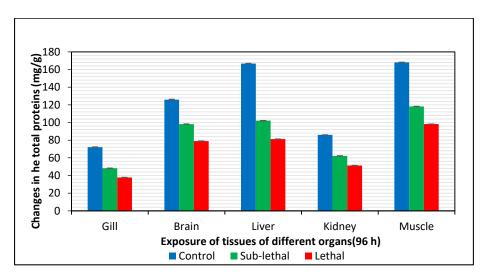


Fig. 4: Changes in protein content (mg/g wet wt of the tissue) in different tissues of fish *Cirrhinus mrigala* (Hamilton) on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50% SP) for 96hours.

Under exposure of sub-lethal and lethal concentrations of Cartap hydrochloride for 24 h, the amount of protein was found to decrease in all the tissue of *Cirrhinus mrigala*.

The lyotropic gradation series in terms of per cent decrement at 24 h and 96 h exposure was:

Sub-lethal -24 h: Liver > Gill > Muscle > Kidney > Brain

Lethal-24 h- Liver > Muscle > Gill > Kidney > Brain

Sub-lethal -96 h: Liver > Gill > Muscle > Kidney > Brain

Lethal-96 h: Liver > Gill > Muscle > Kidney > Brain

For 24 h exposure maximum per cent of the decrease in total protein was observed in the liver (sub-lethal 23.05, lethal 28.33). For 24 h exposure minimum percentage of depletion of total protein was found in the brain (sub-lethal 11.18, lethal

13.13). For 96 h the percentage of decrease was maximum in the liver (sub-lethal 38.72, lethal 51.27). Similarly, for 96 h minimum percentage of decrease was noticed in the brain (sub-lethal 22.06, lethal 37.30).

Protein is the most primary biochemical ingredient present in large quantities in the body of fish. Liver is rich in protein and centre for various metabolism of the fish. In the present study maximum decrease of total protein in the liver is due to the increased rate of proteolytic activity or repeated break down of protein to yield energy due to stress caused due to pesticide exposure. Anitha & Rathnamma (2016) noticed decreased protein levels in all the tissues like liver, kidney, brain, gill and muscle of *Labeo rohita* exposed to lethal and sub-lethal concentrations of Pyraclostrobin 20%

WG (carbamate) for 24 h and sub-lethal concentrations for 5 and 10 days. A decline in the protein content was noticed by Kumari et al. (2014) in the liver of Clarias batrachus when exposed to sub-lethal concentrations (2 and 4 mg. L^{-1}) of the Carbaryl for 96 h. A decrease in protein may be due to the impairment of protein synthesis or an increase in the rate of its degradation to amino acids. Exposure to Carbryl for 4 and 24 days, decreased protein content in liver and muscle of fish Mugil cephalus, when exposed to the lethal and sub-lethal concentration of Carbaryl for 4 days and 21 days respectively was reported by Shivanagouda et al. (2013). Kumar et al. (2017) reported a reduction in proteins in the liver and kidney of freshwater fish, Channa punctatus exposed to different sub-lethal concentrations of pesticide Carbaryl for a period of 15, 30, 45, 60, 75 and up to 90 days. Muddassir (2015) observed significant decrease value in Total protein in the liver of Channa punctatus was treated with 0.1 mL Carbofuran and 0.09 mL Malathion pesticides at different time intervals 7, 14, 21 and 28 days. The decrement of total protein may be due to the inhibition of RNA synthesis disturbing the protein metabolism or this may be due to liver damage where most protein synthesis usually occurs, these results agreed with that of Singh & Sharma (1998). The depletion of protein might also be attributed to spontaneous utilization of amino acids in various catabolic reactions inside the organism to combat the stress condition (Borah & Yadav 1996). Wankhedkar & Bhavsar (2015) found that total protein content significantly decreased in foot and hepatopancreas in land snail C. moussonianus when treated with Cartap hydrochloride and Imidacloprid at lowest concentration i.e. LC₅₀ 0.41ppm and LC₅₀ 0.54 ppm respectively. Veeraiah et al. (2018) observed a decrease in protein content

in all tissues exposed to lethal and sub-lethal concentrations of cyhalothrin 2.5% EC, in the fish *Ctenopharyngdon idellus* for 24 and 96 h.

Proteins are important organic constituents of the animal cells. Understanding the protein components of the cell becomes necessary in the light of the radical changes taking place in protein profiles during pesticide intoxication (Anitha & Rathnamma 2016). The decreased trend of the protein content as observed in the present study in most of the fish tissues may be due to metabolic utilization of the ketoacids through gluconeogenesis pathway for the synthesis of glucose or due to the directing of free amino acids for the synthesis of necessary proteins, or for the maintenance of osmotic and ionic regulation (Schmidt Neilson 1975).

Nucleic Acids (DNA & RNA)

The changes in DNA content observed in the various tissues of *Cirrhinus mrigala* after the Cartap hydrochloride exposure along with the control was graphically represented in Fig. 5 and Fig. 6.

In control fish, DNA content present in different organs was in the order of:

Muscle > Gill > Brain > Liver > Kidney.

Under exposure to sub-lethal and lethal concentrations of Cartap hydrochloride, for 24 h the amount of DNA was found to decrease in all the tissue of *Cirrhinus mrigala*. The lyotropic gradation series in terms of per cent decrement at 24 h and 96 h exposure was:

Sub-lethal -24 h: Gill > Liver > Muscle > Brain > Kidney Lethal-24 h-Gill > Liver > Muscle > Brain > Kidney

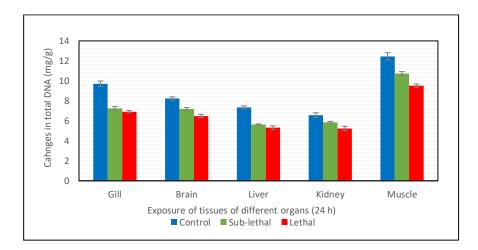


Fig. 5: Changes in the amount of deoxyribonucleic acid (DNA) (mg/g wet weight of the tissue) in different tissues of the fish, *Cirrhinus mrigala* on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50% SP) for 24 h.

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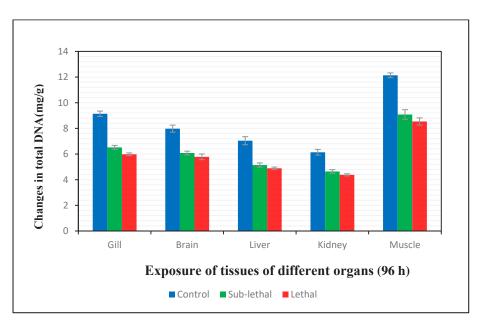


Fig. 6: Changes in the amount of deoxyribonucleic acid (DNA) (mg/g wet wt of the tissue) in different tissues of the fish *Cirrhinus mrigala* on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50% SP) for 96 h.

Sub-lethal-96 h: Gill > Liver > Muscle > Kidney > Brain Lethal-96 h: Gill > Liver > Muscle > Kidney > Brain

In the present study under Cartap hydrochloride exposure for 24 h, maximum percentage of depletion in amount of DNA was noticed in Gill (sub-lethal 25.44, lethal 28.74). Minimum percentage of depletion was exhibited in the Kidney (sub-lethal 10.95, lethal 20.37). Under exposure to sub-lethal and lethal concentrations of Cartap hydrochloride for 96 h, maximum percentage of depletion was seen in Gill (sub-lethal 28.67, lethal 34.58) and minimum percentage of depletion was noticed in the brain (sub-lethal 23.81, lethal 27.57).

The changes in RNA content observed in the various tissues of *Cirrhinus mrigala* after Cartap hydrochloride exposure along with the control was graphically represented

in Fig. 7 and Fig. 8.

The RNA content in different tissues in control fish *Cirrhinus mrigala* was in the order of:

Muscle > Brain > Gill > Liver > Kidney.

Under exposure to lethal and sub-lethal concentrations of Cartap hydrochloride, for 24 and 96 h, the amount of RNA was found to decrease in all the tissues. The lyotropic gradation series in terms of per cent decrement at 24 h and 96 h exposure was:

Sub-lethal -24h: Muscle > Gill > Kidney > Brain > Liver Lethal-24h: Muscle > Gill > Kidney > Liver > Brain Sub-lethal -96h: Gill > Muscle > Kidney > Liver > Brain Lethal-96h: Gill > Muscle > Liver > Kidney > Brain

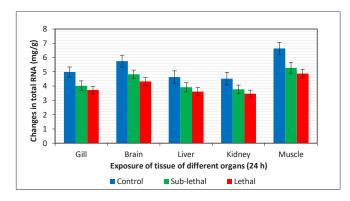


Fig. 7: Changes in the amount of ribonucleic acid (RNA) (mg/g wet wt of the tissue) in different tissues of the fish, *Cirrhinus mrigala* on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50% SP) for 24 h.

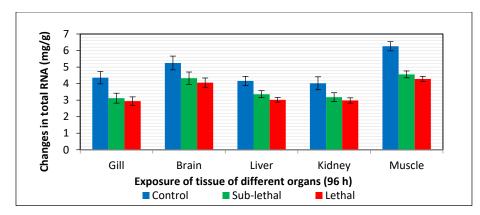


Fig. 8: Changes in the amount of ribonucleic acid (RNA) (mg/gm wet wt of the tissue) in different tissues of the fish, *Cirrhinus mrigala* on exposure to sub-lethal and lethal concentration of Cartap hydrochloride (50% SP) for 96 h.

After 24 h of exposure maximum percentage of decrease in the amount of RNA was found in muscle (sub-lethal 20.48, lethal 26.65). Minimum per cent of the decrease in RNA was found in the liver (15.51) in sub-lethal concentration and brain (19.65) in lethal concentration. After 96h of exposure maximum percentage of decrease in the amount of RNA was found in Gill (sub-lethal 28.44, lethal 32.56). Minimum percentage of decrease in RNA was found in brain(sub-lethal 17.52, lethal 22.66).

The results indicated that the DNA and RNA content in all the tissues of test fish were decreased compared to controls and the decreasing trend was more pronounced in lethal concentrations than in sub-lethal concentrations. In maintaining the physiological configuration of the fish Nucleic acids play a vital role. As Nucleic acid and protein play the main role in regulating different activities of cells they are regarded as important biomarkers of the metabolic potential of cells (Veeriah et al 2013a).

The decrease in nucleic acid content in the present study was in accordance with Vivek (2015) in fingerlings of *Labeo rohita* exposed to sub-lethal concentrations of Cartap hydrochloride for 24, 48, 72 and 96 h. Similar results were also found by Dasu (2014) in fingerlings of *Labeo rohita* were exposed to Thiocarb (Larvin 75% WP) a thiocarbamate pesticide. Anitha & Rathnamma (2016) noticed decreased DNA and RNA levels in all the tissues like liver, kidney, brain, gill and muscle of *Labeo rohita* exposed to lethal and sub-lethal concentrations of Pyraclostrobin 20% WG (carbamate) for 24 h and sub-lethal concentrations for 5 and 10 days.

Tilak et al. (2009) noticed a decreased level of DNA and RNA content in Alachlor treated freshwater fish, *Channa punctatus* (Bloch). The decrease of RNA may be due to inhibiting the function of RNA polymerase or due to interference in the incorporation of precursor in the nucleic acid synthesis. The alterations in DNA levels may be due to disturbances in DNA synthesis and its turnover rate besides degenerative changes caused by pesticides.

From the present study, it can be concluded that exposure of *Cirrhinus mrigala* to Cartap hydrochloride caused a decline in the glycogen, total protein and Nucleic acids (DNA, RNA) content which is more pronounced in lethal exposure than in sub-lethal exposure. The alterations caused during pesticide exposure may be due to the decreased catabolism of the biomolecules to meet the energy demand of test organism under stress or their reduced synthesis due to impaired tissue function. Therefore, the results of this study suggest a serious concern towards the potential danger of Cartap hydrochloride for the aquatic environment and organisms suggesting judicious and careful use of this pesticide in the agricultural area.

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