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Selective Detection of Zn²⁺ lons by Ratiometric Receptor (E)-N'-(1-(2, 5-Dihydroxy phenyl) Ethylidene) Isonicotinohydrazide: A DFT Study

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Detection of zinc Ratiometric receptor Isonicotinohydrazide DFT study

INTRODUCTION

ABSTRACT

The metal ion sensing characteristics of a novel Schiff-based ratiometric UV-visible chemosensor (E)-N'-(1-(2,5-dihydroxy phenyl)ethylidene) isonicotinohydrazide) (R1) has been explored. In EtOH:H₂O (7:3, v/v), it has high sensitivity and selectivity for Zn²⁺ among a series of metal ions. With the addition of Zn²⁺ ions solution, R1 displayed discriminating spectral activity. The other metal ions did not affect R1 in any way. Furthermore, the addition of Zn²⁺ ions to R1 and LMCT action caused the shifting of the peak to a longer wavelength of 406 nm. The interaction of Zn²⁺ ions with R1 was further investigated using Density Functional Theory (DFT) investigations. Zn²⁺-R1 combination has a lower energy (2.2667 kcal.mol⁻¹ to 0.9339 kcal.mol⁻¹) than R1, indicating a strong connection with excellent stability. The Zn²⁺-R1 complex's association constant (Ka) was discovered to be 6795M⁻¹ and 6836M⁻¹ using Benesi-Hildebrand and Scatchard plots respectively. The detection limit was determined to be 276 nM.

Many industrial processes rely heavily on transition metals including iron, zinc, and nickel. For the production of ammonia and fertilizers, iron is employed as a catalyst. Nickel is a key component of various steel-based alloys due to its high strength and resilience to corrosion and heat in the steel industry. Galvanization of iron is generally done with zinc. In the paint and dye-based industries, iron and zinc oxides are extensively employed as pigments (Lehn 1995, Atood et al. 1996).

Zinc is often seen securely bound in proteins in the human body. It plays a range of vital roles in biological processes. Infantile diarrhea, Parkinson's disease, and Alzheimer's disease, immunological dysfunction might all be caused by a disturbance of Zn(II) homeostasis (Lehn 1995, Atood et al. 1996, De Silva et al. 1997, Kumar et. al. 2018, An et al.

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Umesh Fegade: https://orcid.org/0000-0002-2599-668X 2016). As a result, creating probes that can properly detect and understand in vivo Zn(II) biological processes requires the ability to observe Zn(II) concentration in biological systems.

Because of their importance in a range of biological and industrial processes, effective real-time atomic detection of all of these metals is critical. For detecting them, several analytical approaches are available (Zhang et al. 2019a, 2020, Khanna et al. 2019, Ghorai et al. 2019, Yan et al. 2018, Dong et al. 2017). Metal sensing based on optical phenomena is a growing subject of study (Fegade et al. 2014a). Because of its simplicity, ease of use, real-time detection, cheap cost, excellent selectivity, and sensitivity, in the environmental, analytical, and biological realms, this technique has a wide range of applications (Fegade et al. 2014b, 2015a, 2015b, Patil et al. 2014, Kuwar et al. 2013, Tayade et al. 2014, Pawar et al. 2015, Patil et al. 2015, 2014, 2020, Bhosale et al. 2015, Kaur et al. 2017, Kolate et al. 2020, Fegade et al. 2021a, 2021b).

In comparison to fluorescence-based chemosensors, other detection techniques such as AAS, XRF, radioisotopes, ICP-MS (Berrones-Reyes et al. 2019), modified nanotube carbon

electrodes (Khanra et al. 2019), voltammetry (Zhang et al. 2019b), and ion-selective membrane electrodes (Diana et al. 2019, Jung et al. 2019) have a substantial benefit. Fluorescent chemosensors are appealing approaches because of their low cost, high selectivity, sensitivity, and ability to detect in real-time (Ghorai et al. 2019, Yan et al. 2018, Dong et al. 2017, Fegade et al. 2014c, Fegade et al. 2015a, 2015b, Patil et al. 2014,). Chemosensors have played a significant role in gaining a better knowledge of biological processes and their applications in the environment. In recent years, chromogenic zinc sensing molecules have been extensively produced for the goal of monitoring and/or detecting the transient and dynamic distribution of zinc ions inside the cell (Fegade et al. 2014d, 2015a, 2015b, 2020, 2021a, 2021b, Patil et al. 2014, 2015, 2021, Jethave et al. 2018, 2019, 2021, Xu et al. 2018, Yan et al. 2018).

Many transition metals are required for the chemistry of biological systems, with Fe, Co, Cu, and Mo being the most prevalent examples. However, because of the severe toxic consequences, simple, sensitive, selective, and Reliable sensing systems or sensors are required for detecting and eliminating metals from diverse environmental matrices (Lehn 1995, Atood et al. 1996, De Silva et al. 1997, Khan et. al 2013). However, these procedures have significant disadvantages, including a time-consuming sample preparation process, expensive costs, and the need for specialized instrumentation. Furthermore, due to the color change effect visible to the naked eye, identifying trace elements with a colorimetric probe is simple. The response time between the analyte and the probe is relatively short, and apparent color changes allow detection to move from the lab to on-site applications (An et al. 2016, Maniyazagan et al. 2017, Dong et al. 2017, Ghosh et al. 2017, Yang et al. 2019, Naha et al. 2020). In Schiff UV-visible/ fluorescent probes, with metal ions, the C=N bond results in the formation of a stable complex., demonstrating the discriminating spectral activity. As a result, Schiff base probes have attracted the attention of several scientists in the fields of environmental and chemical sciences (Pendin et al. 2019).

For a few years, our team has been working on detecting and removing organic and inorganic contaminants from the environment (Fegade et al. 2014e, 2015a, 2015b, 2020, 2021a, 2021b, Patil et al. 2014, 2015, 2021, Jethave et al. 2018, 2019, 2021). In this study, (E)-N'-(1-(2,5-dihydroxyphenyl) ethylidene) isonicotinohydrazide) (R1) a Schiff base, was produced and tested in fluorescence and UV-visible spectrophotometers for the detection of Zn^{2+} ions. Color shift was observed by the human eye and wavelength enhancement and the shift was examined for the validation of Zn^{2+} interaction with the R1.

MATERIALS AND METHODS

Fisher Scientific Chemicals Ltd. in India provided all reagents, which were utilized without being purified further. The "1H NMR" spectra were acquired on a "Bruker Avance II 400 NMR" operating at 400 MHz in DMSO. The MASS and IR spectra were generated using water "QTOF-Q micro mass" and "Bruker FTIR spectrometers", respectively. A Shimadzu UV-1800 model with 1 cm quartz cells was used to perform the UV-visible spectrophotometric examination.

Synthesis of R1

A solution of isoniazide (0.411 g, 3 mmol) and 2,5-dihydroxy benzaldehyde (0.414 g, 3 mmol) are mixed in 50 mL of CH_3OH and provided 2 h of stirring at room temperature results in the yellow precipitate, which was purified, dehydrated, and recrystallized from ethanol (85%) to yield pure R1 (Fig. 1).

Photographic and Physical Examination

UV-visible spectrophotometer was used for the analysis of working solution R1 to assess metal ion selectivity, then various metal ions were added. R1 demonstrated a discriminating spectrum shift when Zn^{2+} ions were added, but other metal ions did not, resulting in photometric titrations.

DFT Research



Fig. 1: Synthesis of R1.

GAUSSIAN 09 software was applied for all computational calculations. The optimal research was conducted at the B3LYP level, utilizing the Gaussian 09 program's $6-311G^{**}$ basis set (for Zn atom). To establish the nature of the reaction, LUMO and HOMO values were used to monitor the interaction of Zn²⁺ ions with R1.

Calculation of the LOD

The detection limit was calculated using the equation below.

LOD (limit of detection) = 3/S

S = Slope.

RESULTS AND DISCUSSION

Chemosensor R1 was made by condensation reaction between isoniazide and 2,5-dihydroxy benzaldehyde in methanol (Fig. 1) and analyzed using Spectro-analytical methods. The purity of the R1 has been established based on IR, ¹H-NMR, and mass spectroscopic techniques. ¹H-NMR $\begin{array}{l} (\text{CDCl3, 400 MHz, ppm, } \delta): 2.43 \ (\text{s}, 3\text{H}, \text{CH}_3), 6.72\text{-}6.79 \\ (\text{m, 2H, Ar-H), 6.980 \ (\text{s}, 1\text{H}, \text{Ar-H}), 7.82\text{-}7.83 \ (\text{m, 2H}, \text{Ar-H}), 8.76\text{-}8.77 \ (\text{m, 2H, Ar-H}), 8.84 \ (\text{s}, 1\text{H}, \text{-}\text{OH}), 11.49 \\ (\text{s}, 1\text{H}, \text{-}\text{NH}), 12.436 \ (\text{s}, 1\text{H}, \text{-}\text{NH}), 1\text{R} \ (\text{KBr, cm}^{-1}) \upsilon = 668.34 \\ 752.60, 781.20, 831.42, 1149.44, 1208.93, 1292.38, 1504.27 \\ 1533.90, 1665.88, 3452.81, \text{MS} \ (\text{EI}): \text{m/z} = 271.0957 \\ (\text{C}_{14}\text{H}_{13}\text{N}_3\text{O}_3) \ \text{cald} \text{m/z} = 272.55163 \ \text{for} \ \text{C}_{14}\text{H}_{13}\text{N}_3\text{O}_3 \ [39\text{-}43]. \end{array}$

The behavior of R1 (C = 1×10^{-5} M) absorption in the presence of different metal ions (C = 1×10^{-4} M) was investigated using UV-visible spectrophotometric measurements. R1's UV-Visible spectra revealed two peaks at 396 nm, with n \rightarrow П* transitions ascribed to the peak. In addition, the addition of Zn²⁺ ion solution to R1 produced a ratiometric spectrum pattern, but the insertion of different ionic species had no effect. Because of the LMCT action, the addition of Zn²⁺ ion solution to R1 resulted in a red shift of the 396 nm peak to 406 nm, as seen in Fig.2a. The complex bond form between Zn²⁺, OH of R1, and C=N was identified. The redshift is generated by the -OH group being deprotonated,



Fig. 2: a) Metal ion selectivity utilizing R1, Inset Fig. 2a Colour change of R1 (Pale yellow) with the addition Zn^{2+} (Orange) and b) R1 absorption profile with gradual addition of Zn^{2+} ion.

which leads to the formation of an O-Zn covalent bond. The addition of Zn^{2+} to R1 results in not only spectral shifts but there are also chromatic changes from yellow to orange, as measured by colorimetry. The color shift is visible with the naked eye (Fig. 2a Inset), increasing R1 efficiency in today's industrially polluted world (Benesi & Hildebrand 1949, Scatchard 1949, Xu et al. 2018).

Titrations were performed to have a better grasp of occurrence. The spectra of R1 were initially recorded, as shown in Fig, 1b, and then a Zn^{2+} ion solution varying in concentration from 0 to 200 μ L was introduced to R1 (2 mL). The absorbance at 396 nm increases as the amount of Zn^{2+} ions increase, allowing Zn^{2+} ions to be detected in both synthetic and actual samples. The absorption ratiometric response of R1 with Zn^{2+} in the presence of different metal

ions was investigated to evaluate selectivity, as shown in Fig. 3a. The interference study is crucial in the detection of target analytes because interfering ions have a major impact on the method's detection capabilities as well as the detection limit (Fig. 3b).

In the FTIR spectra of pure ligand, broadbands were seen at 3423 cm⁻¹ and 3095 cm⁻¹, which correspond to the –OH and –NH stretching frequencies, respectively. Carbonyl and imine stretching frequency characteristic bands formed sharply at 1668 cm⁻¹ and 1525 cm⁻¹, respectively. In the FTIR spectra of the equivalent R1-Zn²⁺ complex, the unique bands associated with -OH and -NH are diminished/ decreased, and the band for carbonyl and imine stretching is moved to a lower wavelength in the complex at 1664cm⁻¹ and 1517cm⁻¹, respectively (Yan et al. 2018, Meng et al.



Fig. 3: a) Ratiomeric graph of R1 with other metals and b) R1 interference investigations with Zn^{2+} in the existence of some of the other metal cations.

2018). For the imidol structure to become the dominant one during complexation, the ligand's amide functionality must have undergone amido-imidol tautomerism, according to the preceding facts. The band for the carbonyl and imine functional groups also shifts to a lesser wavelength due to the participation of N in the C=N with the Zn. m.z⁻¹ = 335.6534 (C₁₄H₁₂N₃O₃Zn) cald m.z⁻¹ = 336.0901 for (C₂₁H₁₈ClCuN₄O₂).

The highest occupied MO and lowest unoccupied MO energy levels of the title sensor ligand R1 and the corresponding sensor ligand R1+Zn²⁺ are calculated computationally at the B3LYP level using the basis function system 6-311++ G(d,p) to realize the impact of Zn²⁺ binding with the sensor ligand R1 and its electronic properties. 3D plots of the highest occupied MO and lowest unoccupied MO are furnished in Fig. 4 and the energy values are depicted in Table 1. The calculated highest occupied MO and lowest unoccupied MO energies of the sensor ligand R1 are -6.3356 eV and -4.0689 eV, respectively and the resulting bandgap energy (Δ E) of the sensor ligand R1 is 2.2667 eV. Alike, in the sensor ligand R1+Zn²⁺, the HOMO has an

energy of -4.3732 eV, whereas the LUMO has an energy of -3.4393 eV, and the bandgap energy is 0.9339 eV. As can be seen from the figure, the electron density in the highest occupied MOs of sensor ligand R1 is mainly confined to the dihydroxyphenyl structural motif while electron density in the lowest unoccupied MOs of the same is chiefly occupied on the pyridinyl scaffold. Besides, the density of electrons in the highest occupied MOs of sensor ligand R1+Zn²⁺ is primarily located on the dihydroxyphenyl unit akin to the pure sensor ligand R1, however, the density of electrons in the lowest unoccupied MOs of the same is positioned predominantly on the metal and its surroundings such as imine, carbonyl and one of the hydroxyl moieties coordinated with the metal. Precisely, when the highest occupied MOs and lowest unoccupied MOs are concerned, the predominant electron distribution region transfers from one province to another in both cases (Gangatharan et al. 2018, Khanra et al. 2019). On comparing the highest occupied MOs and the lowest unoccupied MOs energy difference between the sensor ligand R1+Zn²⁺ and the sensor ligand R1, the former has a lower value when compared to the latter reflecting the higher stability of the



Fig. 4: DFT calculation (a) R1 and R1. Zn2+, (b) LUMO, (c) HOMO diagrams.

Parameter	Energies (eV)		
	R1	R1+Zn ²⁺	
НОМО	-6.3356	-4.3732	
LUMO	-4.0689	-3.4393	
ΔΕ	2.2667	0.9339	

Table 1: Energies value of S1and Sensor 3+Zn²⁺

sensor ligand R1+ Zn^{2+} . Consequently, the results obtained from computational calculations are in good harmony with

the experimental analysis of the Zn^{2+} binding (Ghorai et al. 2019, Zhang et al. 2019a, 2019b, Awad et al. 2019).

The complex's binding constant may also be calculated by titrating R1 with a Zn^{2+} ion solution. Using Benesi-Hildebrand and Scatchard plots, the association constant (Ka) of the Zn^{2+} -R1 complex was determined to be 6795M⁻¹ and 6836M⁻¹, respectively (Fig. 5a and 5b). As seen in Fig. 6, the binding stoichiometry is a constantly fluctuating process. The absorbance of the host H/([H]+[G]) was also shown in Fig. 6 for a range of total concentrations (Berrones-Reyes





Fig. 5: a) Benesi-Hildebrand for R1 where the K_a value at 6795M⁻¹ and R² = 0.982, and b) Scatchard graph for R1 where the K_a value at 6836 M⁻¹ and R² = 0.9981.



Fig. 6: Job plot between R1 and Zn²⁺ showing 1:1 complexation.

et al. 2019). The concentration of Zn^{2+} and R1 complex was found to be at a maximum of 0.5 units in Fig. 6. 276 nM detection limit (LOD) was found for the 1:1 stoichiometry of Zn^{2+} and R1 (Panunzi et al. 2019).

Reversibility and Colorimetrically Naked Eye Detection

Using absorption spectra, many ions were employed to investigate the reversible binding of Zn^{2+} ions. The brown hue of the Zn^{2+} -R1 was only removed with the addition of $EDTA^{2-}$. It's easy to see how the addition of $EDTA^{2-}$ may shatter the Zn^{2+} -R1 combination. The reversible reagent $EDTA^{2-}$ is used

to demetallize the compound and restore R1 to its natural color. Fig. 7 shows that R1 has greater reversibility of 6 cycles. The resulting complex was extracted with the EDTA²⁻ after the Zn²⁺ ion sensing technique. In comparison to the published sensors (Li et al. 2019, Gu et al. 2019, Xu et al. 2019, Zhang et al. 2019a, 2019b), the regenerated R1 may be used numerous times to better detect Zn²⁺ ions, signifying sensitivity and cost-effectiveness. With the recommended colorimetric R1, the color shift from pale yellow to orange (Fig. 8) may be seen with the naked eye, as a consequence of this a reliable and selective sensor system for in-situ applications was designed (Mao et al. 2013, Guan et al. 2019, Park et al. 2014).



Fig. 7: A Receptor 1 reversible cycle with Zn²⁺ and EDTA²⁻



Fig. 8: a) Reversible cycle of R1 with Zn^{2+} and $EDTA^{2-}$, and b) graph of the reversible study, Inset figure capture of reversibility (R1, R1+Zn and R1+Zn+ EDTA).

ANALYTICAL APPLICATIONS

Recovery tests with real samples were done to assess the proposed colorimetric probe's selectivity, sensitivity, and reliability. Tap water and Mineral samples were taken from the industrial region of Bhusawal, Maharashtra, for the recovery tests. Zn^{2+} ions solutions with concentrations ranging from 10 to 25 g.L⁻¹ were introduced into the water samples. Table 2 showed that utilizing R1 resulted in better Zn^{2+} recoveries (> 97%) and lower percentage relative standard deviation (percent RSD) values (Cox et al. 2019, Bagheri et al. 2020, Zhou et al. 2014, Yang et al. 2019, Pendin et al. 2019, Yang 2019, Huang et al. 2019, Swami et al. 2018, Sargenti et al. 2014, Zhao et al. 2015, Kang et al. 2013).

CONCLUSIONS

Chemosensor R1 was made by condensation reaction between isoniazide and 2,5-dihydroxy benzaldehyde in

methanol and analyzed using Spectro-analytical methods. The UV-Visible spectra of R1 revealed two peaks 396nm and $n \rightarrow \Pi^*$ 396 nm transitions are assigned to the peak. In addition, the addition of Zn^{2+} ion solution to R1 produced a ratiometric spectrum pattern, other metal ions, however, had no effect. Because of the LMCT action, the addition of Zn^{2+} ion solution to R1 resulted in a red shift of the 396nm peak to 406 nm. The Zn^{2+} -R1 complex's association constant (Ka) was discovered to be 6795M⁻¹ and 6836M⁻¹ using Benesi-Hildebrand and Scatchard plots respectively. The 1:1 stoichiometry of Zn^{2+} and R1 having a detection limit (LOD) of 276 nM.

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Sample	Zn^{2+} added (µg.L ⁻¹)	Zn^{2+} found (µg.L ⁻¹) using R1	Recovery (%) by R1 (n=3)
Mineral water	10	9.86 ± 0.045	99.01
	15	13.99 ± 0.084	99.24
	25	24.72 ± 0.053	99.32
Tap water	10	9.89 ± 0.049	98.42
	15	14.71 ± 0.047	98.61
	25	24.71 ± 0.037	98.88

Table 2: Results of a Real sample of Zn²⁺ sensing

N = Number of samples.

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