

Nature Environment and Pollution Technology An International Quarterly Scientific Journal

No. 1

Vol. 8

2009

pp. 33-35

Original Research Paper Effect of Superoxide Dismutase and Catalase in Erythrocytes of Catfish (*Heteropneustes fossilis* Bloch.) Exposed to

# Cadmium

### M. V. Radhakrishnan and S. Hemalatha

Department of Zoology, Annamalai University, Annamalainagar-608 002, T.N., India

Key Words: Heteropneustes fossilis Erythrocytes Superoxide Dismutase Catalase Cadmium exposure

## ABSTRACT

Effect of sublethal concentration of cadmium chloride (12 ppm; 10% of 96h  $LC_{50}$ ) in erythrocytes of catfish was studied for a period of 5, 10, 20 and 45 days. It was found that the activity of superoxide dismutase (SOD) in red blood cells (RBC) significantly decreased after 5 and 10 days of cadmium exposure. However, the SOD activity increased after 20 and 45 days of cadmium treatment. Elevated activity of catalase (CAT) was found in erythrocytes of cadmium treated fish after 45 days. These results indicate oxidative stress and cell damage in the exposed fish.

### INTRODUCTION

In freshwater fishes cadmium enter through gills and binds to albumins and erythrocytes in the blood, and then transferred into tissues and organs where it is bound to proteins of low molecular mass producing metallothioneins by the induction of metallothionein mRNA synthesis (Gould & Karolus 1974, George et al. 1996). It causes significant metabolic alterations and injuries at different levels (Pratap & Bonga 1990, Brown et al. 1984). The synthesis of metallothioneins in certain organs decreases to some extent the toxicity of non-bound cadmium (Thornalley & Vasak 1985, Wormser et al. 1990, Olsson & Kille 1997). In fish, cadmium causes destruction of erythrocytes and leads to anaemia (Pushpa 2008). Cadmium increases the production of reactive oxygen species (ROS) in tissues and inhibits the activity of some enzymes of the antioxidative defense system (Jackim et al. 1970, Pruell & Engelhardt 1980, Zikic et al. 1996, 2001). There are only scarce data available on influence of cadmium on the activity of enzymes of the antioxidative defense system in erythrocytes of fishes (Thomas & Wofford 1993, Palace et al. 1993, Zikic et al. 1997). Hence, the present investigation has been designed to understand the influence of cadmium on the activity of superoxide dismutase (SOD, EC 1, 15. 1.1) and catalase (CAT, EC 1.11.1.6) in erythrocytes of catfish (*Heteropneustes fossilis*) after exposure lasting 5, 10, 20 and 45 days.

## MATERIALS AND METHODS

The experimental catfish (*Heteropneustes fossilis* Bloch.) weighing 82-85 g were adapted for 30 days to the laboratory conditions with water temperature of  $28 \pm 2^{\circ}$ C, pH 7.4 and concentration of dissolved oxygen 5.6 ppm (APHA 1998) dechlorinated and aerated water. The fish were fed with minced goat liver. After the period of acclimation, four experimental groups of fish were exposed to cadmium in a concentration of 12 ppm in water (10% of 96 h LC<sub>50</sub> = 120 ppm) (Hamilton et al. 1977). Control fish were resided in non-polluted water. The fish were sacrificed in groups after exposure to cadmium for 5, 10, 20 and 45 days, each group consisting of 10 fish.

Table 1: The activity of superoxide dismutase (SOD) and catalase (CAT) in erythrocytes of control catfish (C) and catfish exposed to 12 ppm cadmium for 5, 10, 20 and 45 days.

Parameters	Experimental				
	Control	5 d	10 d	20 d	45 d
SOD	430 ± 1.12	$352 \pm 0.75*$	410 ± 1.15	598 ± 2.10*	$657 \pm 1.05*$
CAT	$1.75\pm0.75$	$1.60\pm0.25$	$1.42\pm0.25$	$1.60\pm0.75$	$1.68\pm0.00$

Note: The values are expressed in units of enzyme activities per mL of red blood cells (U/mL RBC). Mean  $\pm$  S.E.M from seven fish in each group. Significantly different from controls; \*p < 0.05.

After sacrificing the fish, freshly heparinised blood samples were collected immediately and prepared for further processing as recommended by Mazeud et al. (1979) and Wdzieczak et al. (1982). The activity of superoxide dismutase (SOD) was determined spectrophotometrically at 480 nm by the epinephrine method, and it was expressed in units of enzyme activity per mL of red blood cells (U/mL RBC). The activity of catalase (CAT) was determined spectrophotometrically at 570 nm (Sinha 1972), and it was expressed in mmoles of decomposed hydrogen peroxide per second per mL RBC (mmol H<sub>2</sub>O<sub>2</sub>/sec/mL/RBC). Data were analysed using the non-parametric Mann-Whitney two-tailed test and differences at p < 0.05 were considered as significant.

#### **RESULTS AND DISCUSSION**

The results are represented in Table 1. Cadmium causes a significant decrease of SOD activity (p < 0.05) after 5 days of exposure. No significant differences were found after 10 days of exposure compared to the control values, whereas after 20 and 45 days of exposure, values were significantly increased (p < 0.05) when exposed in U/mL RBC. The obtained data show that cadmium did not change the activity of CAT expressed in mmol  $H_2O_2$ /sec/mL RBC.

In the present study, the activity of SOD was significantly decreased after 5 days of exposure to sublethal cadmium. Similar results were obtained in previous investigations (Zikic et al. 1997) on carp during acute cadmium exposure. However, after 10 days, the activity of this enzyme was non-significant with control values. During prolonged exposure, the activity of SOD was significantly increased after 20 and 45 days. These results indicate the activation of protective mechanisms necessary for scavenging of produced free radicals of oxygen in erythrocytes. Kostic et al. (1993), while studying the effect of cadmium on antioxidant and metabolic status in RBC of rats, showed the enhanced activity of this enzyme. In the present study cadmium did not exhibit any effects on the activity of CAT when expressed per mL RBC. Petronijevic et al. (1995) observed significant alterations of energy metabolism in RBC when rats exposed to cadmium. It also increased concentration of non-enzymatic components of the antioxidant defense system in plasma (ascorbic acid-AsA, Vitamin E) (Zikic et al. 1995) of rats and enhanced activity of enzymes of antioxidant defense system SOD, CAT, glutathione peroxidase and glutathione reductase in RBC of rats (Kostic et al. 1993).

#### ACKNOWLEDGEMENT

The authors are grateful to Dr. M. Sabesan, Professor and Wing Head, Department of Zoology, for providing the laboratory facilities and encouragement.

#### REFERENCES

APHA 1998. Standard Methods for the Examination of Water and Wastewater, 20th edn. American Public Health Associa-

Vol. 8, No.1, 2009 • Nature Environment and Pollution Technology

tion, Washington DC., USA.

- Brown, D.A., Bay, S.M., Alfara, J.F., Hershelman, G.P. and Rosenthal, K.D. 1984. Detoxication/toxification of cadmium in scorpion fish (*Scorpena guttata*) acute exposure. Aquat. Toxicol., 5: 93-107.
- George, S.G., Todd, K. and Wright, J. 1996. Regulation of metallothionein in teleosts-induction of MT mRNA and protein by cadmium in hepatic and extrahepatic tissue of a marine flatfish, the turbot (*Scophthalmus maximus*). Comp. Biochem. Physiol., 113: 109-115.
- Gould, E. and Karolus, J.J. 1974. Physiological response of the cunner, Tautogolabrus disperses to cadmium. NOA Tech. Rep., NMFS, SSRF, 681: 21-25.
- Hamilton, M.A., Russo, R.C. and Thurston, R.V. 1977. Trimmed spearman Karber method for estimating median lethal concentrations in toxicity bioassays. Environ. Sci. Technol., 11: 714-719.
- Jackim, E., Hamlin, J.M. and Sonis, S. 1970. Effects of metal poisoning on five liver enzymes in the killifish (Fundulus heteroclitus). J. Fish. Res. Board. Can., 27: 383-390.
- Kostic, M.M., Ognjanovic, B., Dimitrijevic, S., Zikic, R.V., Stajn, A., Rosic, G.L. and Ivkovic, R.V. 1993. Cadmium induced changes of antioxidant and metabolic status in red blood cells of rats *in vivo* effects. Eur. J. Haematol., 51: 86-92.
- Mazeud, F., Maral, J. and Michelson, A.M. 1979. Distribution of superoxide dismutase and glutathione peroxidase in the crop: erythrocyte manganese SOD. Biochem. Biophys. Res. Commun., 86: 1161-1168.
- Olsson, P.E. and Kille, P. 1997. Functional comparison of the metal-regulated transcriptional control regions of metallothionein genes from cadmium-sensitive and tolerant fish species. Biochem. Biophys. Acta, 1350: 325-334.
- Palace, V.P., Majewski, H.S. and Klavercamp, J.F. 1993. Interactions among antioxidant defences in liver of rainbow trout (Onchorhyncus mykiss) exposed to cadmium. Can. J. Fish. Aquat. Sci., 50: 156-162.
- Petronigivic, M.R., Maletic, S.D., Zikic, R.V. and Kostic, M.M. 1995. Glycolysis and oxidative pentose phosphate pathway in red blood cells of rats chronically intoxicated with cadmium. Coll. Sci. Pap. Fac. Sci. Krag., 17: 191-201.
- Pratap, H. and Bonga, S.E.W. 1990. Effects of water-borne cadmium of plasma cortisol and glucose in the cichlid fish Oreochromis mossambicus. Comp. Biochem. Physiol. C., 95: 313-317.
- Pruell, R.J. and Engelhardt, F.R. 1980. Liver cadmium uptake, catalase inhibition and cadmium thionein production in the killifish (*Fundulus heteroclitus*) induced by experimental cadmium exposure. Mar. Environ. Res., 3: 101-111.
- Pushpa, 2008. Cadmium chloride induced alterations in haematological parameters of the freshwater fish *Channa striatus* (Bloch.). M.Phil. Thesis, Annamalai University, India.
- Sinha, A.K. 1972. Colorimetric assay of catalase. Anal. Biochem., 47: 389-394.
- Thomas, P. and Wofford, H.W. 1993. Effects of cadmium and Arochlor-1254 on lipid peroxidation, glutathione peroxidase activity and selected antioxidants in Atlantic croaker tissues. Aquat. Toxicol., 27: 159-178.
- Thornalley, P.T. and Vasak, M. 1985. Possible role of metallothionein in protection against radiation-induced oxidative stress. Kinetics mechanism of its reaction with superoxide and hydroxyl radicals. Biochem. Biophys. Acta, 827: 36-44.
- Wdzieczak, J., Zalesna, G., Wujec, E. and Peres, G. 1982. Comparative studies on superoxide dismutase, catalase and peroxidase levels in erythrocytes and livers of different freshwater and marine fish species. Comp. Biochem. Physiol. B. 73: 361-365.
- Wormser, U., Benzakine, S. and Nyska, A. 1990. Cadmium induced metallothionein synthesis in the rat liver slice system. Toxicol. in vitro, 4: 791-794.
- Zikic, R.V., Skajn, A., Saicic, Z.S. Spasic, M.B., Ziemnicki, K. and Petrovic, V.M. 1996. The activities of superoxide dismutase, catalase and ascorbic acid content in the liver of goldfish (*Carasiusauratus gibelio* Bloch.) exposed to cadmium. Physiol. Res., 45: 479-481.
- Zikic, R.V., Stajn, A., Ognjanovic, B., Pavlovic, S.Z. and Kostic, M.M. 1995. The effects of cadmium and selenium on the ascorbic acid and vitamin E contents in the plasma and liver of young and adult rats. Coll. Sci. Pap. Sci. Krag., 17: 203-213.
- Zikic, R.V., Stajn, A.S., Ognjanovic, B.I., Pavlovic, S.Z. and Saicic, Z.S. 1997. Activities of superoxide dismutase and catalase in erythrocytes and transaminases in the plasma of crops (*Cyprinus carpio* L.) exposed to cadmium. Physiol. Res., 46: 391-396.