

Nature Environment and Pollution Technology An International Quarterly Scientific Journal

ABSTRACT

Vol. 10

Twenty seven fungal species were found associated with seeds of six cereals. Maximum fungi were reported from seeds of *Pennisetum typhoides* and *Sorghum vulgare*. Aspergillus flavus, Fusarium moniliformae and

Fusarium oxysporum were found pathogenic causing seed rot, seed discoloration and germination inhibition.

2011

pp. 273-275

Original Research Paper

# Fungal Biodiversity in Seeds of Some Cereals of Nashik District, and Its Pathogenicity and Control Measures

Captan and Dithane M-45 proved best for seed dressing.

#### D. N. Khairnar, A. S. Kelhe and A. B. Khairnar

Mycology and Plant Pathology Research Laboratory, Department of Botany, K. A. A. N. M. S. Arts, Commerce & Science College, Satana-423 301, Maharashtra, India

Nat. Env. & Poll. Tech. Website: www.neptjournal.com

Received: 17-9-2010 Accepted: 27-10-2010

Key Words: Seed mycoflora, Cereals Pathogenicity

## INTRODUCTION

The seed-borne fungi of cereals were earlier studied by Sharma & Basu Chaudhary (1975), Gupta (1976), Konde et al. (1880), Randhwa & Aulakh (1980), Prasad & Narayan (1981), Girisham & Redddy (1985, 1986) and Khairnar & Mukadam (1989).

The present investigations were carried out to detect the seed-borne fungi of some major cereals viz., *Pennisetum typhoides, Sorghum vulgare, Triticum aestivum, Zea mays, Oryza sativa* and *Elsusine coracana* by different seed health testing methods and to study their pathogenic behaviour and control by seed dressing fungicides.

# MATERIALS AND METHODS

Seed samples of bajra (pearl millet), jowar, wheat, maize, rice and elsusine were collected in three random samples (half kg each) from fields, various storehouses and markets. A composite sample of this was prepared by mixing the individual samples, and preserved in cloth bags at laboratory temperature during the study.

Standard blotter and agar plate method with Wakman's acid agar and Rose bengal agar medium was used as recommended by ISTA (1960) for the isolation of seed-borne fungi of six cereals. Four hundred seeds were used in each case. Seeds used for experiments were untreated and pretreated with 0.1%  $HgCl_2$  solution; in agar plate method ten seeds were plated in each plate. The plates were incubated at  $20 \pm 2^{\circ}C$  under alternate light and darkness condition for seven days.

The pathogenicity tests of each fungus on seeds during germination were carried out by soaking the surface sterilized seeds in spore suspensions of seed-borne fungi for 24 h. The seeds were used for germination studies on moist blotter. Seeds treated similarly but without spore suspension served as control. This type of work was done by Panchal (1984) on jowar seeds.

The fungicides namely Captan, Dithane M-45, Dithane Z-78, Brassicol, Blitox-50 W, Bavistin, Thiram, Zinkop, Ceresan, Zineb-75, Wettable sulphur each (2 g/kg seed) were evaluated for their efficacy in reducing the seed-borne fungi of pearl millet. The treated seeds were tested by standard blotter method after 24 hours of the treatment. Untreated seeds served as control.

#### **RESULTS AND DISCUSSION**

It is clear from the results summarized in Table 1 that 27 fungal species appeared on the seeds of six cereals tested. In present investigation three fungi viz., *Mortirella exigua*, *Pythium* sp. and *Torula herbarum* are newly recorded on *Pennisetum typhoides*. On untreated seeds, maximum incidence was of *Drechslera tetramera* followed by *Aspergillus niger*, *A. flavus*, *Fusarium oxysporum*, *F. moniliformae* and *Rhizopus nigricans*, while *Absidia ramosa*, *A. flavipes*, *A. fumigatus*, *Drechslera rostrata*, *Penicillium oxalicum*, *Pithium* sp., *Torula herbarum* and *Rhizoctonia solani* were reported poorly.

Seeds treated with surface sterilizer showed complete absence of certain fungi like *Absidia ramosa*, *Aspergillus nidulans* and *Penicillium oxalicum*. It was interesting to note that one phycomycetous non-sporulating fungus appeared on untreated seeds. Fungal species like *Aspergillus flavus*, *A. niger* and *Fusarium oxysporum* were found on all cereals.

It is evident from the results given in Table 2 that

	Fungal species	% incidence on the seeds											
Sr. No.		Pennisetum typhoides		Sorghum vulgare		Triticum aestivum		Zea mays		Oryza sativa		Elsusine coracana	
		UT	РТ	UT	PT	UT	PT	UT	РТ	UT	РТ	UT	PT
1.	Absidia ramosa	10	_	10	-	-	-	-	-	-	-	-	-
2.	Alternaria alternata	-	10	20	10	-	-	-	-	-	-	-	-
3.	Aspergillus carbonarius	20	10	30	20	20	20	-	-	10	-	-	-
4.	A. flavipes	10	-	10	10	10	-	-		-	-	-	-
5.	A. flavus	30	20	30	10	30	10	40	10	30	20	40	10
6.	A. fumigatus	10	-	-	-	-	-	-	-	-	-	-	-
7.	A. nidulans	10	-	10	-	20	-	10	-	10	-	-	-
8.	A. niger	40	30	20	20	20	10	10	10	20	20	30	-
9.	A. terreus	10	10	-	-			-	-	-	-	-	-
10.	A. ustus	20	10	10	10	20	10	20	-	-	-	10	-
11.	Cladosporium herbarum	20	10	20	10	10	-	-	-	-	-	-	-
12.	Curvularia lunata	10	10	20	20	-	-	-	-	-	-	-	-
13.	C. Pallescens	-	20	10	-	-	10	-	-	-	-	-	-
14.	Drechslera longirostrata	20	30	20	20	-	-	-	-	-	20	-	-
15.	D. rostrata	10	10	10	10	-	-	-	10	-	-	-	-
16.	D. tetramera	60	10	10	10	-	-	-	-	-	-	-	-
17.	Fusarium moniliformae	30	10	20	20	-	10	-	-	-	-	-	-
18.	F. oxysporum	30	20	10	20	40	-	30	20	-	10	-	10
19.	Mortierella exigua	20	10	10	-	-	-	-	-	-	-	-	-
20.	Mucor globosum	20	20	20	10	30	-	40	10	20	-	-	-
21.	Penicillium oxalicum	10	-	10	-	-	-	-	-	-	-	-	-
22.	Pythium sp.	-	10	-	-	-	-	-	-	-	-	-	-
23.	Rhizoctonia solani	-	10	10	10	-	-	-	-	-	-	-	-
24.	Rhizopus nigricans	30	30	30	-	30	-	-	-	-	-	-	-
25.	Syncephalastrum racemosum	20	-	20	10	-	-	-	-	-	-	-	-
26.	Torula herbarum	10	10	-	-	-	-	-	-	-	-	-	-
27.	Non-sporulating mycelium	10	-	-	-	-	10	-	-	-	-	-	-
	Total fungial species $= 27$	23	20	21	16	10	07	06	05	05	04	03	02

Table: 1. Fungal biodiversity in seeds of some cereals of Nashik district.

UT = Untreated seeds; PT = Pretreated seeds; + Present; - Absent

Table 2: Effect of artificial infestation on seeds and seedlings of Pennisetum typhoides.

	Abnormalities in seeds and seedlings									
Fungal species	% Seed	Seed	Seed Discolourations	Seedlin	ng	Seedling				
	Germination	Rot	Discolourations	Shoot	Length	Root	Length			
Alternaria alternata	40	-	Black brown	Blight	5.0	-	9.8			
A. tenuis	40	-	Brown	Yellow	5.2	Shortening	4.1			
Aspergillus flavus	10	+	Green	Tip rot	2.6	Root rot	1.9			
Aspergillus niger	100	-	black	Yellow	5.4	Healthy	9.0			
Cladosporium herbarum	40	-	Dull green	Stunted	1.6	Shortening	3.0			
Curvularia lunata	30	-	Black cloudy	Chlorosis	4.9	Root rot	10.7			
Curvularia pallescens	50	-	Black cloudy	Stunted	2.5	Root rot	9.2			
Dreshslera longirostrata	40	-	Black cloudy	Blight	5.0	Root rot	10.0			
Dreshslera rostrata	50	-	Black cloudy	Blight	5.2	Root rot	9.2			
Dreshslera tetramera	20	-	Black cloudy	Blight	5.0	Root rot	8.8			
Fusarium moniliformae	00	+	White pink	-	-	-	-			
Fusarium oxysporum	00	+	White	-	-	-	-			
Penicillium oxalicum	10	-	Faint blue	Pale green	2.8	Curling root	4.5			
Rhizopus nigricans	20	-	Ash	Tip rot	4.5	Shortening	1.5			
Rhizoctoni solani	50	-	Black brown	Chlorossis	4.9	Root rot	9.7			
Control	100	-	Normal	Normal	5.2	Normal	10.0			

Vol. 10, No. 2, 2011 • Nature Environment and Pollution Technology

274

complete inhibition of seed germination was achieved due to *Fusarium moniliformae* and *Fusarium oxysporum*, while seed rotting was effectively found due to *Aspergillus flavus*, *Fusarium moniliformae*, *F. oxysporum*, and partial seed rot by *Penicillium oxalicum*. In five days old seedlings, blight and retardation of root length and shoot elongation were the common symptoms caused by the most of the seed-borne fungi.

Captan, Dithane-45, Bavistin and Blitox-50W (each 2g/kg seeds) showed broad spectrum effect and eliminated all the fungi from seed and improved germination to the extent of 90-98 percent as compared to 50-60 percent obtained in untreated seeds. The remaining fungicides were less effective in controlling the seed-borne fungi of cereals.

## ACKNOWLEDGEMENT

Authors are thankful to UGC, New Delhi for providing financial assistance for this work and also thankful to the Principal of the college for providing necessary facilities and encouragement during the work.

# REFERENCES

- Girisham, S. and Reddy, S.M. 1985. Influence of storage structures on seed mycoflora of pearl millet. Geobios New Reports, 4: 126-129.
- Girisham, S. and Reddy, S.M. 1986. Seed mycoflora of pearl millet (*Pennisetum americanum*) in relation to seed moisture. Geobios New Reports, 5: 121-125.
- Gupta, D.C. 1976. Viability of stored seeds in bajra (*Pennisetum typhoides*). Seed Tech. News, 6: 9-12.
- ISTA 1966. Proc. Int. Seed Test Ass., 31: 1-52.
- Khairnar, D.N. and Mukadam, D.S. 1989. Seed moulds of pearl millet, their pathogenicity and control. Geobios New Reports, 8: 41-44.
- Konde, B.K., Dhage, B.V. and More, B.B. 1980. Seed-borne fungi of some pearl millet cultivars. Seed Research, 8: 59-63.
- Panchal, V.H. 1984. Studies on seed-borne fungi of Sorghum. Ph.D. Thesis, Marathwada University, Aurangabad.
- Prasad, B.K. and Narayan, N. 1981. Seed-borne fungi of some millets. Geobios, 8: 47-48.
- Randhawa, H.S. and Aulakh, K.S. 1980. Pathological appraisal of seedborne fungi of pearl millet. Indian Phytopathol., 33: 163-167.
- Sharma, J.R. and Basuchaudhary, K.C. 1975. Assessment of seed mycoflora of pearl millet and their control. Indian Phytopathol., 28: 388-390.